

# Extraction of 45 Multi Class Drugs of Abuse from Urine (Non-Hydrolyzed) Using ISOLUTE® SLE+ prior to LC-MS/MS Analysis

This application note describes a Supported Liquid Extraction (SLE) protocol for the extraction of various drugs of abuse from non-hydrolyzed urine prior to LC-MS/MS analysis.

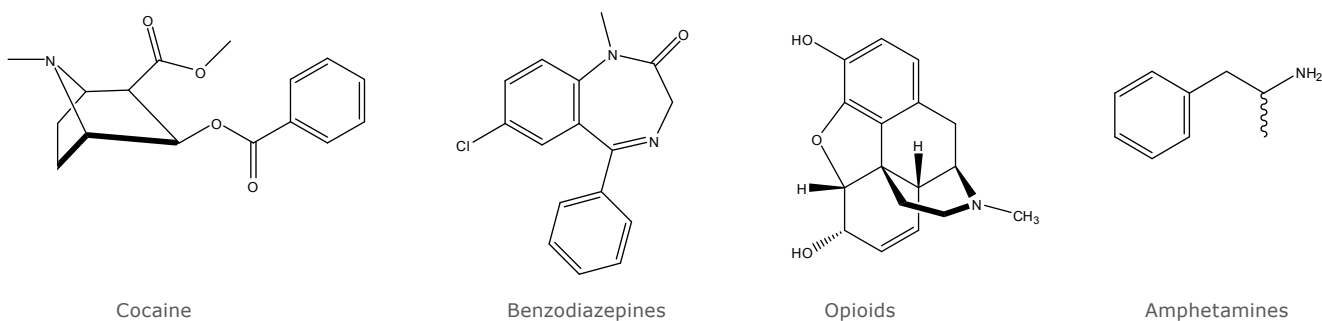


Figure 1. Example structures by class

## Introduction

The method described in this application achieves high reproducible recoveries for a wide range of drugs of abuse and metabolites from non-hydrolyzed urine. Urine was extracted using the ISOLUTE SLE+ 200 fixed well plate and 1 mL column formats using 100 and 500  $\mu$ L sample volumes, respectively. Sub ng/mL levels were achieved for all analytes using 100  $\mu$ L of urine on the fixed well plate format.

ISOLUTE® SLE+ products provide clean, rapid, robust, efficient, high throughput and automatable extraction solutions for a wide range of analytes.

## Analytes

Amphetamine, Methamphetamine, MDA, MDMA, MDEA, Methadone, EDDP, Mephedrone, Morphine, Hydromorphone, Oxymorphone, Dihydrocodeine, Oxycodone, Hydrocodone, Codeine, 6-MAM, Cocaine, Benzoyllecgonine, Fentanyl, Norfentanyl, Ketamine, Norketamine, Buprenorphine, Norbuprenorphine, 7-amino-flunitrazepam, 7-amino-clonazepam, Nitrazepam, Flunitrazepam, Clonazepam,  $\alpha$ -OH-alprazolam,  $\alpha$ -OH-triazolam, Oxazepam, Estazolam, Temazepam, Alprazolam, Lorazepam, 2-OH-ethyl-flurazepam, Triazolam, Nordiazepam, Diazepam, Midazolam, Flurazepam, Zaleplone, Zopiclone, Zolpidem.

## Sample Preparation Procedure

<b>SLE+ Format</b>	ISOLUTE® SLE+ 200 µL sample volume supported liquid extraction plate, part number 820-0200-P01 or ISOLUTE® SLE+ 1 mL sample volume supported liquid extraction columns, part number 820-0140-C.
<b>Urine Pre-Treatment</b>	Take 1 mL urine and spike internal standard (10 µL of Amphetamine-d3, Morphine-d3, Diazepam-d5, parent concentration 250 ng/mL; BZE-d3 and 6-MAM-d3, metabolite concentration 25 ng/mL). Add 1 mL of 1% ammonium hydroxide (aq, v/v) and vortex mix thoroughly

## Supported Liquid Extraction

<b>Sample Loading</b>	<b>ISOLUTE SLE+ 200 µL plate:</b> Load pre-treated sample (200 µL) to the 96 well-plate followed by a pulse of vacuum or positive pressure to initiate flow. Leave to absorb for 5 minutes.
	<b>ISOLUTE SLE+ 1 mL columns:</b> Load pre-treated sample (1 mL) to the columns followed by a pulse of vacuum or positive pressure to initiate flow. Leave to absorb for 5 minutes.
<b>Elution</b>	<b>ISOLUTE SLE+ 200 µL plate:</b> Apply 1 mL of dichloromethane/isopropanol (95/5, v/v) and allow to flow under gravity for 5 minutes. Pull through the remaining solvent with vacuum or positive pressure for 10-20 seconds.
	<b>ISOLUTE SLE+ 1 mL columns:</b> Apply 2.5 mL of dichloromethane/isopropanol (95/5, v/v) and allow to flow under gravity for 5 minutes.
	Apply a second 2.5 mL aliquot of dichloromethane/isopropanol (95/5, v/v) and allow to flow under gravity for 5 minutes. Pull through the remaining solvent with vacuum or positive pressure for 10–20 seconds.
	Note: The addition of 100 µL of 50 mM HCl in methanol into the collection plate or culture tube is required prior to or post elution to stabilize multiple analytes (amphetamines, bath salts and ketamine) due to volatility issues during evaporation.
<b>Post Elution</b>	Evaporate to dryness at 40 °C in a stream of air or nitrogen using a SPE Dry.
<b>Reconstitution</b>	<b>ISOLUTE SLE+ 200 µL plate:</b> 200µL 80/20 2 mM NH <sub>4</sub> OAc H <sub>2</sub> O/MeOH. Cap with a sealing mat and vortex gently.
	<b>ISOLUTE SLE+ 1 mL columns:</b> 500 µL 80/20 2 mM NH <sub>4</sub> OAc H <sub>2</sub> O/MeOH and vortex gently.

## UPLC Conditions

<b>Instrument</b>	Waters ACQUITY UPLC 20 $\mu$ L Loop
<b>Column</b>	ACQUITY UPLC BEH C18 column (1.7 $\mu$ m, 100 x 2.1 mm id)
<b>Mobile Phase</b>	A: 2 mM ammonium acetate (aq) B: 2 mM ammonium acetate in methanol
<b>Flow Rate</b>	0.3 mL/min

**Table 1.** Gradient Conditions

Time	% A	% B	Curve
0	90	10	1
10	10	90	6
11.4	10	90	6
13.4	90	10	1

**Curve 1.** Conditions in line initiated immediately once previous time passed. i.e. 90:10 resumed at 11.4 minutes.

**Curve 6.** Linear Gradient

<b>Injection Volume</b>	10 $\mu$ L (partial loop with overfill)
<b>Sample Temperature</b>	20 °C
<b>Column Temperature</b>	40 °C

## Mass Spectrometry Conditions

<b>Instrument</b>	Waters Premier XE triple quadrupole mass spectrometer equipped with an electrospray interface for mass analysis.
<b>Desolvation Temperature</b>	450 °C
<b>Ion Source Temperature</b>	120 °C

Positive ions acquired in the multiple reaction monitoring (MRM) mode:

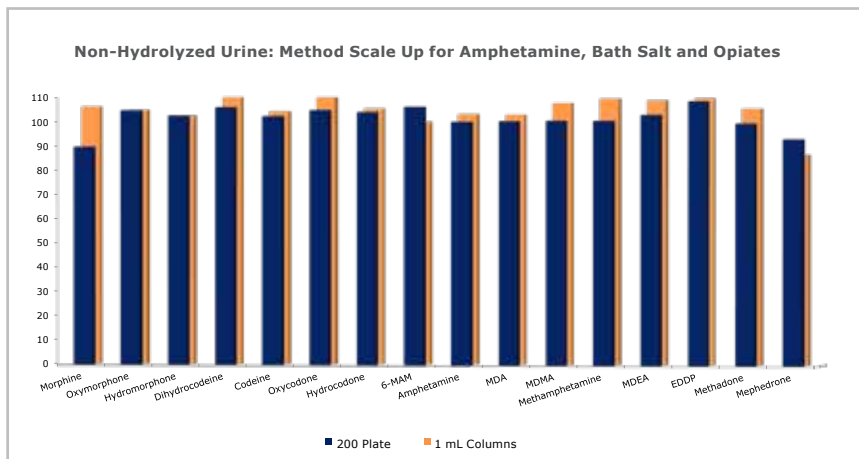
Table 2. MRM Conditions

Compound	MRM Transition	Cone Voltage (V)	Collision Energy (eV)
Amphetamine	136.0 > 118.9	16	9
Amphetamine-d5	141.0 > 123.9	16	9
Methamphetamine	150.0 > 90.9	22	17
MDA	180.1 > 105.0	16	23
MDMA	194.1 > 163.0	20	13
MDEA	208.2 > 163.0	22	13
Hydromorphone	286.2 > 185.1	44	29
Morphine	286.2 > 201.0	42	25
Morphine-d3	289.2 > 201.0	42	25
BZE	290.1 > 168.0	30	18
BZE-d3	293.1 > 171.0	30	18
Oxymorphone	302.2 > 198.1	34	37
Dihydrocodeine	302.2 > 199.1	42	33
Oxycodone	316.2 > 241.2	34	27
Mephedrone	178.1 > 160.0	35	12
Norfentanyl	233.1 > 84.0	25	19
7-amino-flunitrazepam	284.2 > 135.0	40	27
7-amino-clonazepam	286.2 > 121.0	40	30
Hydrocodone	300.2 > 199.1	46	33
Codeine	300.3 > 215.1	42	25
6-MAM	328.2 > 165.1	44	33
6-MAM-d3	331.2 > 165.1	44	33
Cocaine	304.2 > 182.0	30	20
Norketamine	224.1 > 124.9	20	23
EDDP	278.2 > 234.2	26	30
Zaleplone	306.2 > 264.2	40	22
Zopiclone	389.2 > 245.1	20	17
Norbuprenorphine	414.3 > 101.0	55	42
Ketamine	238.1 > 124.9	25	27
Nitrazepam	282.2 > 236.1	40	25
Flunitrazepam	314.2 > 268.2	40	25
Clonazepam	316.1 > 270.1	40	25
$\alpha$ -OH-triazolam	359.1 > 331.1	45	26
Oxazepam	287.2 > 241.0	30	21
Estazolam	295.2 > 267.2	40	24
Temazepam	301.1 > 255.1	30	22
Zolpidem	308.2 > 235.1	45	35
Alprazolam	309.2 > 281.2	40	26
Methadone	310.2 > 265.2	26	15
Lorazepam	321.1 > 275.1	30	22
$\alpha$ -OH-alprazolam	325.2 > 297.1	40	25
2-OH-ethyl-flurazepam	333.2 > 109.0	40	27
Triazolam	343.0 > 308.1	45	27
Nordiazepam	271.1 > 139.9	40	28
Diazepam	285.2 > 154.0	40	27
Diazepam-d5	290.2 > 154.0	40	27
Midazolam	326.2 > 291.2	45	29
Fentanyl	337.3 > 105.0	35	40
Flurazepam	388.2 > 315.1	35	23
Buprenorphine	468.3 > 101.0	55	42

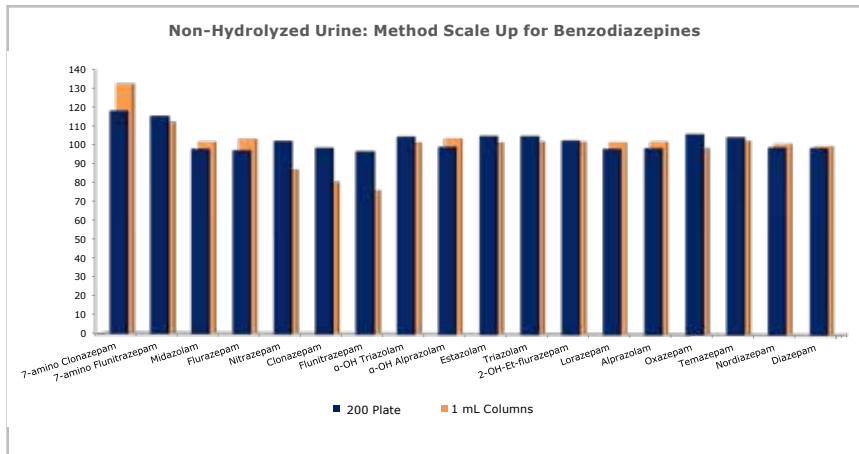
## Results

### Recovery

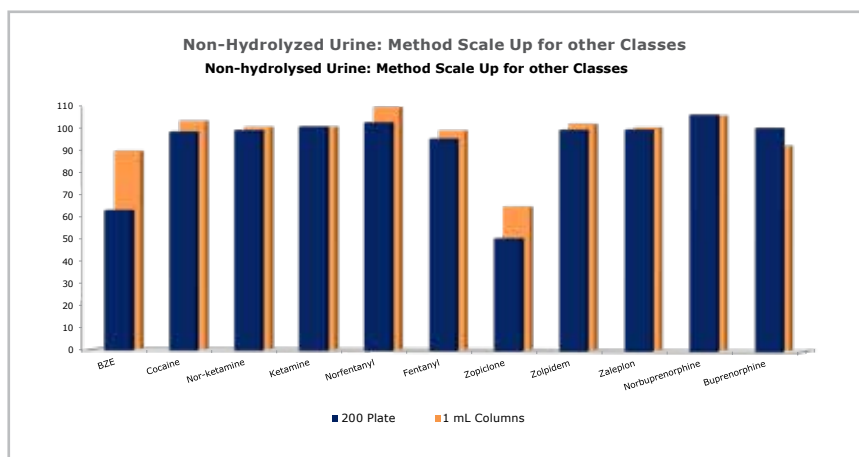
Urine spiked with 2 ng of analytes (n=7), equating to 20 ng/mL or 4 ng/mL when extracting 100 or 500  $\mu$ L of urine, respectively. RSD's (N=7) ranged from 0.8–6.5%.



**Figure 2.** Recovery profile for amphetamines, bath salts and opiates from non-hydrolyzed urine using ISOLUTE® SLE+ 200 fixed well plates and 1 mL columns.



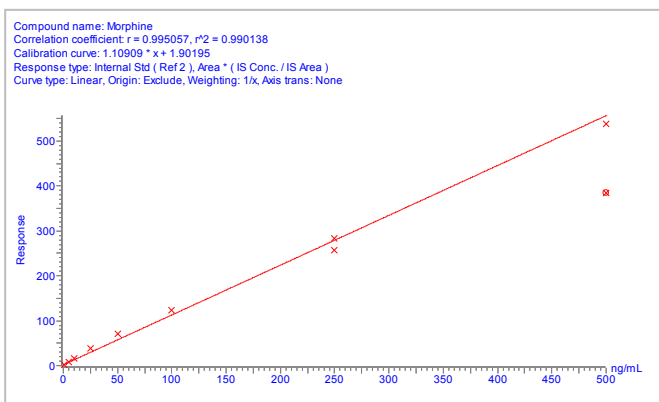
**Figure 3.** Recovery profile for benzodiazepines from non-hydrolysed urine using ISOLUTE® SLE+ 200 fixed well plates and 1 mL columns.



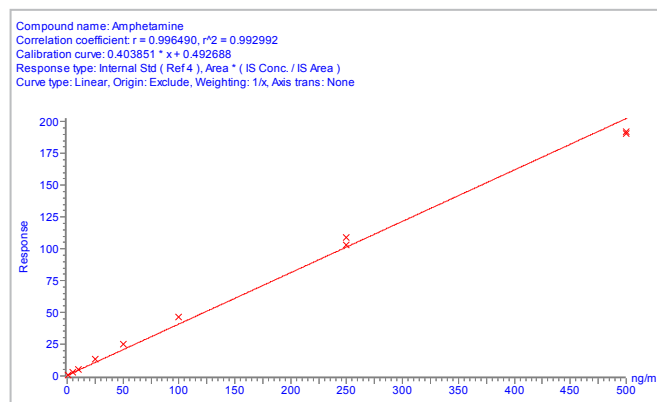
**Figure 4.** Recovery profile for multi class analytes from hydrolyzed urine using ISOLUTE® SLE+ 200 fixed well plates and 1 mL columns.

## Calibration Curves

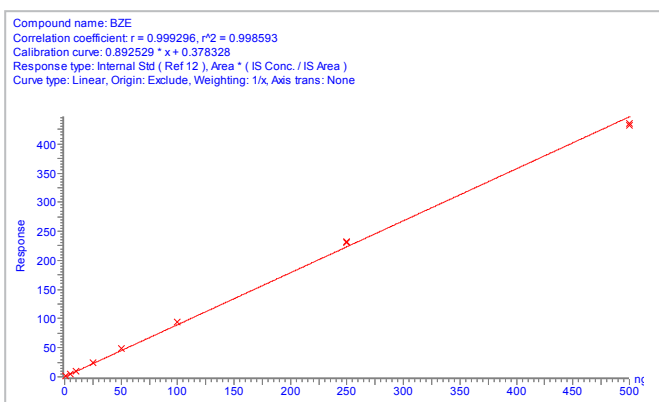
200  $\mu$ L fixed well plate and 1 mL column processing: Calibration curves were generated using urine spiked at concentrations from 1–500 ng/mL. Good coefficients of determination were obtained for all analytes ( $r^2 > 0.99$ ). Quadratic function was observed at the top end of the calibration curve for many analytes. Dilution of these samples was performed to improve linearity.



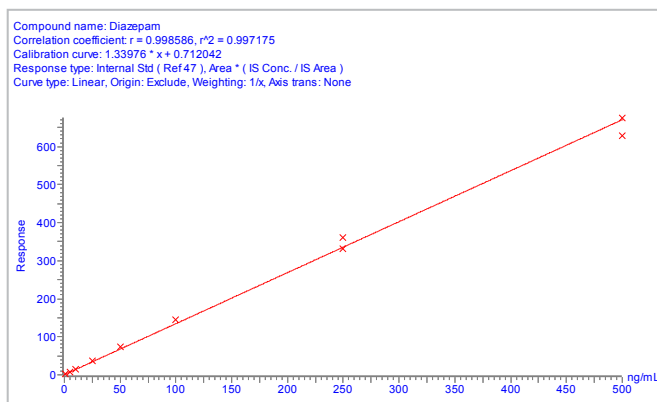
**Figure 5.** Curve for morphine using ISOLUTE® SLE+ 1 mL columns.



**Figure 6.** Calibration Curve for amphetamine using ISOLUTE® SLE+ 1 mL columns.



**Figure 7.** Calibration Curve for benzoylcegonine (BZE) using ISOLUTE® SLE+ 1 mL columns.



**Figure 8.** Calibration Curve for diazepam using ISOLUTE SLE+ 1 mL columns.

**Table 3.** Estimated LOQs based on S/N ratios from 1 ng/mL extracted samples are:

Analyte	Estimated LOQ 200 µL Plate ng/mL	Estimated LOQ 1 mL Columns ng/mL
Amphetamine	< 0.3	< 0.2
Methamphetamine	< 0.15	< 0.1
MDA	< 0.2	< 0.1
MDMA	< 0.1	< 0.05
MDEA	< 0.2	< 0.1
Hydromorphone	< 0.2	< 0.2
Morphine	< 0.5	< 0.2
BZE	< 0.1	< 0.05
Oxymorphone	< 0.5	< 0.5
Dihydrocodeine	< 0.1	< 0.1
Oxycodone	< 0.5	< 0.5
Mephedrone	< 0.5	< 0.25
Norfentanyl	< 0.1	< 0.05
7-amino-flunitrazepam	< 0.2	< 0.1
7-amino-clonazepam	< 0.2	< 0.1
Hydrocodone	< 0.25	< 0.2
Codeine	< 0.25	< 0.1
6-MAM	< 0.25	< 0.2
Cocaine	< 0.1	< 0.05
Norketamine	< 0.2	< 0.1
EDDP	< 0.15	< 0.1
Zaleplone	< 0.2	< 0.1
Zopiclone	< 0.1	< 0.1
Norbuprenorphine	< 0.5	< 0.4
Ketamine	< 0.05	< 0.05
Nitrazepam	< 0.25	< 0.1
Flunitrazepam	< 0.2	< 0.1
Clonazepam	< 0.25	< 0.1
α-OH-triazolam	< 0.5	< 0.2
Oxazepam	< 0.2	< 0.2
Estazolam	< 0.2	< 0.1
Temazepam	< 0.1	< 0.1
Zolpidem	< 0.1	< 0.05
Alprazolam	< 0.15	< 0.1
Methadone	< 0.15	< 0.1
Lorazepam	< 0.75	< 0.4
α-OH-alprazolam	< 0.5	< 0.2
2-OH-ethyl-flurazepam	< 0.5	< 0.2
Triazolam	< 0.2	< 0.1
Nordiazepam	< 0.2	< 0.1
Diazepam	< 0.2	< 0.1
Midazolam	< 0.15	< 0.1
Fentanyl	< 0.15	< 0.1
Flurazepam	< 0.2	< 0.1
Buprenorphine	< 0.5	< 0.3

## Additional Notes

### Buffer Preparation

- 1% ammonium hydroxide aq (v/v): Take 99 mL of H<sub>2</sub>O and add 1 mL of NH<sub>4</sub>OH (28-30% stock concentration).
- 2 mM ammonium acetate aq: Weigh 0.15416 g and dissolve in H<sub>2</sub>O. Dilute and make up to 1 L in H<sub>2</sub>O.
- 2 mM ammonium acetate in methanol: Weigh 0.15416 g and dissolve in H<sub>2</sub>O. Dilute and make up to 1 L in MeOH.
- Dichloromethane/isopropanol (95/5, v/v): Take 95 mL of dichloromethane and add 5 mL of isopropanol
- 80/20 2 mM NH<sub>4</sub>OAc H<sub>2</sub>O/MeOH Reconstitution solvent: Take 80 mL of 2 mM ammonium acetate (aq) and add 20 mL of 2 mM ammonium acetate in methanol

### Blowdown Stability

Amphetamines, bath salts and ketamines suffer blow down issues when drying in the free base form. To combat this effect we added 100 µL of 50 mM HCl in MeOH to the collection plate/culture tubes to convert to the corresponding HCl salt forms.

**Reconstitution volumes can be reduced in order to reach lower limits of quantitation, if required!**

## Ordering Information

Part Number	Description	Quantity
820-0200-P01	ISOLUTE SLE+ 200 Supported Liquid Extraction Plate	1
820-0140-C	ISOLUTE SLE+ 1 mL Supported Liquid Extraction Columns	30
121-9600	VacMaster-96 Sample Processing Manifold	1
PPM-96	Biotage PRESSURE+ 96 Positive Pressure Manifold 96 Well.	1
SD-9600-DHS	SPE Dry sample evaporator	1
C103264	TurboVap 96	1

For the latest application notes and more information about ISOLUTE® SLE+ visit [www.biotage.com](http://www.biotage.com)

#### EUROPE

Main Office: +46 18 565900  
Toll Free: +800 18 565710  
Fax: +46 18 591922  
Order Tel: +46 18 565710  
Order Fax: +46 18 565705  
[order@biotage.com](mailto:order@biotage.com)

#### NORTH AMERICA

Main Office: +1 704 654 4900  
Toll Free: +1 800 446 4752  
Fax: +1 704 654 4917  
Order Tel: +1 704 654 4900  
Order Fax: +1 434 296 8217  
[ordermailbox@biotage.com](mailto:ordermailbox@biotage.com)

#### JAPAN

Tel: +81 3 5627 3123  
Fax: +81 3 5627 3121  
[jp\\_order@biotage.com](mailto:jp_order@biotage.com)

#### CHINA

Tel: +86 21 2898 6655  
Fax: +86 21 2898 6153  
[cn\\_order@biotage.com](mailto:cn_order@biotage.com)

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