A SIMPLE AND SENSITIVE METHOD FOR THERAPEUTIC MONITORING OF AMISULPRIDE IN HUMAN PLASMA/SERUM

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Introduction

Amisulpride (Solian, Sanofi-Synthelabo) is a second generation(atypical) antipsychotic. It is a dopamine antagonist with high selectivity for dopamine D_2 and D_3 receptors [1]. Low dose (50-300 mg/d) treatment is used in patients with predominantly negative symptoms of schizophrenia when it seems to facilitate dopaminergic transmission [1,2]. For patients suffering from acute episodes of schizophrenia with predominantly positive symptoms or from chronic schizophrenia with positive and negative symptoms, higher doses (400-1200 mg/d) are given [1,3]. Corresponding to the the two dose ranges, two plasma threshold concentrations associated with improved treatment efficacy (28-92 μ g/L at low-dose and >153 μ g/L at high dose) have been suggested [4]. The two main metabolites of amisulpride are both inactive, with much of the drug being excreted unchanged in urine.

We have developed a simple and sensitive method for the measurement of total amisulpride [(-)S- and (+)R- amisulpride] in human plasma/serum. The limit of accurate measurement is 2 μ g/L with a 200 μ L sample.

Materials and methods

Amisulpride was a gift from Sanofi-Synthelabo (UK). The internal standard (benzimidazole), Tris (tris[hydroxymethyllaminomethane) and newborn calf serum were from Sigma-Aldrich, and ammonium perrelhorate was from Fluka. Coarsely filtered human serum was from Scipac (Sittingbourne, Kent). Methanol and methyl text-butyl ether (MTBE) were HPLC grade. Water was deionised.

HPLC system: PU-1580 intelligent pump, AS-950 autosampler, and FP-1520 fluorescence detector (excitation: 270 nm, emission: 350 nm) (all Jasco, UK), and Atlas chromatography data system (Thermo, UK). The pre-column and analytical column, stainless steel tubes (10 and 125 x 4.6 mm i.d., respectively) contained Waters Spherisorb® S5SCX sulphopropyl-modified silica (5 μm aps) (both Hichrom, UK). The eluent was methanol containing 0.2 % perchloric acid (70 %) (flow-rate 1.0 mL/min).

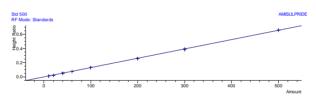
Calibration solutions were prepared in newborn calf serum by appropriate dilution of stock amisulpride solution (100 mg/L free base in methanol) to give concentrations of 10, 20, 40, 60, 100, 150, 200, 300, 500 µg/L amisulpride. Internal quality control (IQC) solutions were similarly prepared in human serum by dilution of separate stock solution to give 25, 75 and 250 µg/L amisulpride. Calibration standards and IQCs were stored at $-20^{\circ}\mathrm{C}$.

Sample preparation : Internal standard solution, aqueous benzimidazole (2 mg/L, 25 $\mu L)$ and Tris buffer (2 mol/L, pH 10.6, 100 $\mu L)$ were added to plasma/serum (200 $\mu L)$. After extraction into MTBE (600 $\mu L)$ by mixing (30 s) and centrifugation (1000 g, 4 min), the extract was evaporated to dryness and the residue reconstituted in methanol. A portion (100 $\mu L)$ of the methanolic extract was analyzed.

Results and Discussion

Using the conditions described, the calibration graph was linear over the calibration range (r > 0.99). The amisulpride response was in fact linear up to 2000 ug/L.

Typical calibration graph for amisulpride



Intra- and inter-assay precision (RSD) were less than 10 % when assessed at three different concentrations (n = 6). Individual batch analyses were accepted if the IQC results were within + 10 % of the nominal values, or + 15 % for the lowest IQC.

Intra-assay precision at three concentrations (n=6)

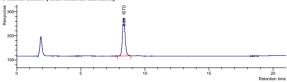
Nominal (µg/L)	Measured (μg/L)	RSD (%)
25	24	9.9
75	63	7.2
250	246	3.6

Inter-assay precision and accuracy at three concentrations (n=6)

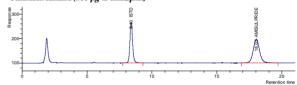
Nominal (μg/L)	Measured (µg/L)	RSD (%)	Accuracy (%)
25	25	3.2	101
75	64	4.1	99
250	240	4.9	96

The extraction of amisulpride from calf serum and human plasma/serum showed no significant difference (paired t-test, p>0.05). In clinical samples (n = 23, median amisulpride dose 1200 mg/d, range 200-1200 mg/d) the median (10^{th} and 90^{th} percentiles) amisulpride concentration was 340 (127-892) $\mu g/L$. Representative chromatograms are shown below:

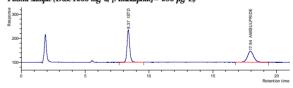




Calibration standard (200 µg/L amisulpride)



Patient sample (Dose 1200 mg/d; [Amisulpride]= 290 µg/L)



Conclusions

The method described is simple, economical, relatively rapid, requires a small sample volume. No inteference from commonly co-prescribed psychoactive drugs has been encountered. The method has sufficient sensitivity to measure amisulpride in plasma's erum from patients prescribed low doses of the drug.

References

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